**International Journal of Food Science and Nutrition** 

ISSN: 2455-4898

Impact Factor: RJIF 5.14 www.foodsciencejournal.com

Volume 2; Issue 6; November 2017; Page No. 83-90



# Effect of high fat diet on appetite regulation and obesity: Endocannabinoid system pathway

# \* Ismail Mucahit Alptekın, Alev Keser

Department of Nutrition and Dietetics, Ankara University, Ankara, Turkey

## **Abstract**

Obesity is an public health problem leading to other metabolic diseases and its prevalance globally increasing. A disturpt in molecular regulation of appetite metabolism causes to obesity by leading to hyperphagy. Dietary pattern affects directly to appetite regulation. Therfore, dietary pattern is so important for obesity prevalance. There are many factor in appetite mechanism having complex system. One of these factors is endocannabinoid system. Over activation of endocannabinoid system relating to dietary fat pattern leads to obesity by increasing food intake. The aim of this research is to investigate effects of dietary fat pattern on obesity through endocannabinoid system.

Keywords: endocannabinoid system, obesity, nutrition, diet

#### 1. Introduction

Obesity is an important public health problem reaching epidemic levels in all age groups around the world [1, 2]. Along with obesity, many metabolic diseases can occur, especially [3] Therefore. cardiovascular diseases interest pathophysiological mechanisms underlying relationship between obesity and metabolic syndrome, diabetes and cardiovascular risk and new therapeutic targets are increasing. Current researchs show that risk factors of obesity alone/in combination play role in risk development and prognosis of disease [4-6]. However, in recent years it has been implicated signaling system a intracellular known endocannabinoid system (ECS) plays an important role in regulation of energy balance, feeding behavior, hepatic lipogenesis and possibly regulation of glucose homeostasis. Since discovery of energy intake regulation and metabolism of ECS, some question markings for pathogenesis of obesity have been removed. Scientific evidence suggests that ECS plays an important role in development of obesity and ECS is overactive in obesity [7-9]. Moreover, it was observed that animals without cannabinoid type 1 (CB1) receptor playing role in appetite regulating and body weight were found to be weak and resistant to diet-related obesity and dyslipidemia [10]. Clinical trials examining pharmacological blockade of CB1 with rimonabant (20 mg/day) in the mid-2000s, this approach has potential to affect many cardiometabolic risk factors (fasting plasma glucose, dyslipidemia) along with abdominal obesity [11-13]. Depending on side effects such as nausea, vomiting, diarrhea, headache, dizziness, anxiety and mood changes [13], Food and Drug Administration (FDA) prevented rimonabant using in obesity treatment in 2007. In 2008, European Medicines Agency (EMEA) has stated that benefits of rimonabant are no more than risks and suggest that it should not be offered for sale [14]. These results make it necessary to clarify mechanisms underlying of factors causing development of obesity and to find more reliable and effective methods in prevention and treatment of obesity. For this reason, in this review, it is aimed to discuss based on evidence role of ECS on obesity development and effect of dietary pattern on ECS.

#### 2. Endocannabinoid System

Endocannabinoid system was identified at the beginning of 1990's with discovery of  $\Delta 9$ -tetrahydrocannabinol (THC) found in *Canabis sativa* [15]. It is a physiological system activated by activation of CB1 and CB2 receptors with anandamide (AEA) and 2-arachidonyl glycerol (2-AG) endocannabinoids [16]. Relationship between ECS and some metabolic diseases, mainly obesity, depends on function of endocannabinoids and cannabinoid receptors.

## 2.1. Endocannabinoids

Endocannabinoids are endogenous fatty acid derivatives can bind to cannabinoid receptors (17). To date, along with AEA and 2-AG, of functions are less known N-arachidonyl dopamine (NADA), 2-arachidonyl glycerol ether (noladine ether) and O-arachidonyl ethanolamine (viroamine) endocannabinoids have been described [18] (Figure 1).

**Fig 1:** Biochemical structures of Anandamide (a) and 2-Arachidonyl glycerol (b) [16].

AEA and 2-AG are composed of arachidonic acid (AA) being intermediate product of linoleic acid metabolism and precursor of other regulatory lipids  $^{[19]}$ . Linoleic acid can easily converted to AA after conversion to  $\gamma$ -linoleic acid, and eicosatetraenoic acid, respectively  $^{[20]}$ . AEA is formed by addition of ethanolamide and phosphatidyl ethanolamide structures to AA accompanied with FAAH enzyme or by conversion of AA to N-arachidonoyl phosphatidyl ethanolamide accompanied with N-acyl transferase enzyme

and Ca<sup>2+</sup> [21]. It is thought that NAPE is the most important resource in AEA formation [22]. 2-AG is formed by conversion of diacylglycerol structure of AA to 2-arachidonyl glycerol with diacylglycerol lipase or by conversion of

phosphatidylinositol with phospholipase A1 <sup>[23]</sup>. The required enzyme is phospholipase C for conversion of phosphatidylinositol bisphosphate to diacylglycerol and lysophosphatidylinositol to 2-AG <sup>[24]</sup> (Figure 2).

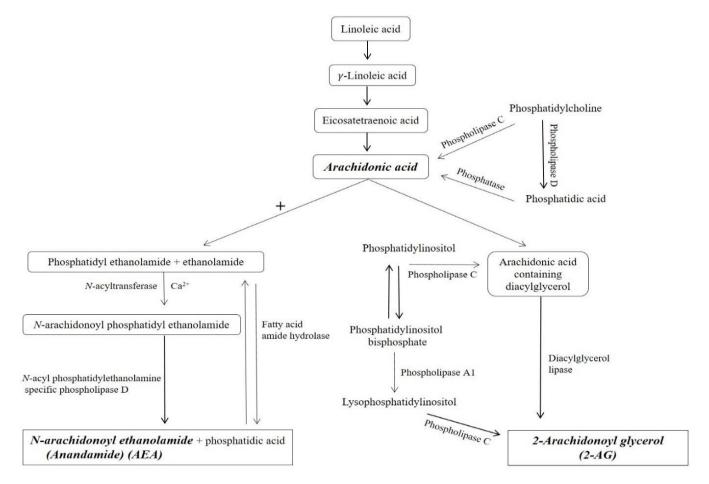


Fig 2: Main pathway of endocannabinoids biosynthesis (AEA, 2-AG) (adapted from Naughton et al. [25]).

Following identification of cannabinoids, cannabinoid receptors and endocannabinoids, effects of ECS on metabolic diseases, mainly obesity, have been investigated <sup>[26, 27]</sup>. As a result of studies, ECS has been shown to regulate food intake and energy balance, to have effects on appetite metabolism and to increase food intake <sup>[28, 29]</sup>.

## 2.2. Endocannabinoid system, appetite and obesity

Hypothalamus plays an important role in regulation of important mechanisms such as energy balance, appetite, hunger-satiety [30]. Following food consumption, hypothalamus is stimulated by hormones secreted by small intestine and central nervous system signals, and satiety occurs. A disturbance in these signals or mechanisms is associated with obesity [31]. It may be evidence that later satisfied and/or to consume more food of obese than normal weight individuals.

Many orexigenic/anorexigenic hormones affect to hypothalamus. In these hormones, it is thought that ghrelin and leptin have role in mechanism of ECS on obesity [32, 33]. Moreover cholecystokinin (CCK) being small intestinal

hormone and has an important role in suppressing appetite, has been shown to reduce expression of cannabinoid receptors stimulating food intake [34].

In a study by Rigamonti et al. [35] examined role of gastrointestinal orexigenic-anorexigenic peptides endocannabinoids on desire to consume chocolate in obese. There was significant correlation between in ghrelin, an organsigenic hormone, and AEA/2-AG levels with chocolate consumption, but wasn't in Peptide tyrosine-tyrosine (PYY) and Glucagon-like peptide-1 (GLP-1) are anorexigenic hormones. It has been concluded that ghrelin and chemical signals such as AEA and 2-AG endocannabinoids are important in terms of nutrition and obesity and that presence of other agents antagonizing effects of these agents may be important in treatment of obesity. These results show an interesting mutual action that CCK, a satiety factor, blocks action of endocannabinoids are orexigenic [36]. Also, activation of ghrelin receptor prevented downregulation of CB1 receptors by CCK, thereby limiting extent of its action [37]. Appetite regulation and food intake control is a complex

Appetite regulation and food intake control is a complex process involving neurological mechanisms [38]. ECS is

thought playing role on obesity by acting on appetite and hunger-satiety mechanisms of hypothalamus causing stimulate food intake [39]. Presence of AEA, 2-AG and CB1 receptors sensing these compounds in hypothalamic areas may be evidence this stuation. In a study on rats, it was found that injection of 50mg AEA into ventromedial hypothalamus resulted in a significant increasing in food intake by hyperphagy development. It may be from appetite stimulating action of AEA is part of stimulation of reward/rewardingbehaviour [40]. In another study, paraventricular hypothalamic nukleus was activated after 45 minutes injection of AEA [41]. Other studies also support this data [42-44]. There are studies of similar results on humans [45-47]. As a result, it is revealed that endocannabinoids increase food intake by activating CB1 receptors in hypothalamus and thus causing obesity development.

# 2.3. Relationship between endocannabinoid system, adipose tissue and lipogenesis

In addition to effects of ECS on appetite metabolism, studies have also been carried out to show that it is effective in some mechanisms particularly in adipose tissue and lipogenesis [48-51]. As a result of increased endocannabinoid levels, canabinoid receptors in adipose tissue are stimulated, lipid metabolism can be affected by various mechanisms, lipogenesis is increased and fatty acid  $\beta$ -oxidation is decreased [52]. In realization of these mechanisms, decreased production of adiponectin [8], increased lipoprotein lipase enzyme activity [48], inhibition of AMP-activated protein kinase enzyme [53] play role.

In a study of presence of endocannabinoids in adipose tissue of healthy subjects, it was found that adipose tissue can bind AEA and 2-AG, which are effective in energy balance and body weight management, and endocannabinoids can play an important role in control of adipose tissue fat mass <sup>[54]</sup>. In another study, level of AA, endocanabinoid precursor, was positively correlated with body mass index (BMI) and obesity <sup>[55]</sup>. In another study, it has been found that there is a relationship between small intestine microbiota and ECS and this plays an important role in control of adipogenesis in intestinal system and adipose tissue <sup>[56]</sup>. These results suggest that endocannabinoids may play an important role in medical treatment of obesity.

Another mechanism, thought ECS causing obesity, is to increase lipogenesis. CB1 receptor activation is known to increase lipogenesis-induced lipid accumulation in both liver [49,57] and fat cells [48, 58, 59]. Bluher *et al.* [51] investigated relationship between circulating endocannabinoid levels and adipose tissue in obese and normal body weight individuals. It was determined that 2-AG levels in circulation were correlated with body fat-mass, visceral fat-mass and fasting

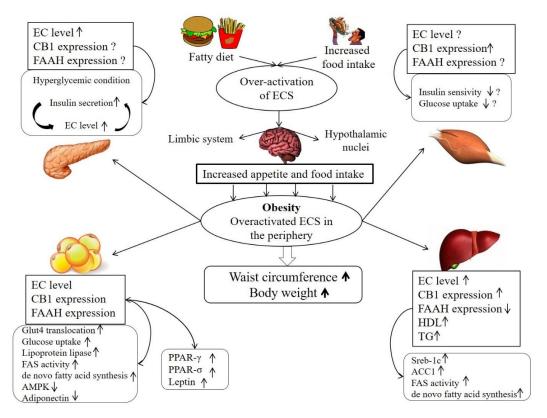
plasma insulin (p <0.05). Moreover, there is a positive relationship between accumulation of abdominal fat and ECS in obese. As a result, it has been emphasized that there must be effective measures for inhibition of ECS in treatment and prevention of abdominal obesity and related metabolic diseases.

In a study by Annuzzi et al. [60] of obese with Type 2 Diabetes, obese without diabetes, and normal weight without diabetes, subcutaneous adipose tissue and AEA, 2-AG, OEA, PEA levels were investigated. It has been stated that AEA concentrations in SAT are associated with increased lipogenesis in liver and this can not be excluded from certain metabolic disorders in obese with diabetes. In conclusion that may be due to overactivation of CB1 receptors stimulated by increased AEA concentrations in SAT. Similar results were found in a study and it was found that endocannabinoid system effects energy balance and causes obesity by stimulating lipogenesis in peripheral tissues and central orexigenic pathway. Cannabinoid receptor antagonists in ECS have been shown to play an important role in prevention and treatment of obesity due to suppression of food intake and lipogenetic pathway [48].

## 2.4. Effects of high fat diet on endocannaniboid system

Eating behavior is regulated by two different systems, homeostatic and hedonic. The homeostatic pathway provides control of energy balance by increasing desire to eat when energy stores are empty. The hedonic pathway is a system based on award increasing desire to delicious food by surpassing homeostatic pathway when energy is excessive in body [61]. Some foods (rich in sugar and/or fat), just as addictives, have been shown to cause increased gene expression especially in nucleus accumbens and increased expression of dinofrinin, an endogenous opioid in arcuate nucleus [62]. Opioid and dopaminergic system are also effective in food enjoyment. It is known that high fat diet (HFD) increases desire to eat by stimulating hedenoic center thereby causing weight gain by distorting energy balance in positive direction [62, 63], it is expressed that HFD also increases body weight through ECS.

HFD stimulates cannabinoid receptors by increasing endocannabinoid levels. Thus, ECS becomes active and causes to increase food intake by stimulating hypothalamic regions controlling hunger-satiety and appetite mechanisms [33, 64, 65]. Given relationship between increasing food intake and increasing obesity risk (66), it can be said that ECS has an important role on development of obesity. ECS also play a role in development of obesity and other metabolic diseases by stimulating adipogenesis and lipogenesis in peripheral tissues [67, 28] (Figure 3). Therefore, endocannabinoids are thought to be a new biomarker for obesity [68].



**Fig 3:** Relationship between endocannabinoid system, food intake and obesity (adapted from Di Marzo and Matias <sup>[65]</sup>, Eckardt *et al.* <sup>[69]</sup>, Vettor and Pagano, 2009 <sup>[70]</sup>). ACC1: Acetyl co-enzyme-A carboxylase-1; AMPK: AMP-activated protein kinase; CB1: Cannabinoid1; EC: Endocannabinoid; ECS: Endocannabinoid system; FAAH: Fatty acid amide hydrolase; FAS: Fatty acid synthase; Glut-4; Glucose transporter type-4; HDL; High density lipoprotein: Sreb-1c: Lipogenic transcription factor (Sterol regulatory element binding transcription factor-1); TG; triglyceride.

Since endocannabinoids are fatty acid derivatives, they are directly related to dietary pattern. Therefore, HFD may lead to the development of obesity through ECS by increasing

endocannabinoid levels. Some studies examining relationship between HFD and ECS shown in Table 1.

Table 1: Effects of dietary pattern on endocannabinoids

Study-Year	Materials-Methods	Results
Engeli <i>et al</i> . 2014 <sup>[71]</sup>	Two weeks isocaloric LFD and HFD in obese and normal weight subjects.	Weight-neutral changes in dietary fat intake cannot explain excessive endocannabinoid availability in human obesity. Obesity and dietary fat intake affect ECS gene expression in a tissue specific manner.
Alvheim <i>et al</i> . 2013 <sup>[72]</sup>	Atlantic salmon were fed FO and SO for 6 months.  Male C57BL/6J mice were fed diets of 35% of energy as fat based on FO- and SO-enriched salmon for 16 weeks.	Excessive dietary LA elevates endocannabinoids in the liver of salmon and mice, and increases weight gain and counteracts the anti-inflammatory properties of EPA and DHA in mice.
Crespillo <i>et al</i> . 2011 [73]	Rats were fed with two different types of diets for 12 weeks: an STD (10% fat) and an HFD (60% fat) in order to induce obesity.	The accumulated caloric intake was progressively higher in rats fed on the HFD than the STD, resulting in a divergence in body weight gain.
Alvheim <i>et al</i> . 2012 [74]	Mice were fed diets containing 1 en% LA, 8 en% LA, and 8 en% LA + 1 en% EPA+DHA in medium-fat diets (35 en% fat) and HFD (60 en%) for 14 weeks from weaning.	Dietary LA increased tissue AA, and subsequently elevated 2-AG+1-AG and AEA resulting in the development of diet-induced obesity.
Martin <i>et al</i> . 2017 <sup>[75]</sup>	Mice were fed for an additional 12 weeks the same phytol-free, phytoestrogen-free, 10 kcal% fat control chow or pair-fed isocaloric HFD.	LKO markedly diminished the impact of HFD on brain endocannabinoid levels.
Batetta <i>et al</i> . 2009 <sup>[76]</sup>	Male Zucker rats were fed for 4 weeks a control diet or diets supplemented with either FO or KO.	Diets rich in (n-3) LCPUFA, and a KO-based diet which was associated with lower endocannabinoid concentrations in several peripheral tissues.
Matias <i>et al</i> . 2008 [ <sup>77</sup> ]	Animals were fed for 14 weeks different HFDs are HFD1; 25.5% fat (49% of calories), 22% protein and 38.4% carbohydrate. HFD2; 33.8% fat (59.9% of	Statistically significant elevations (in the skeletal muscle, heart and kidney) or reductions (in the thyroid) of the levels of either AEA or 2-AG, or

	calories), 23.9% protein and 27.1% carbohydrate.	both, were found.
Piscitelli <i>et al</i> . 2011 <sup>[78]</sup>	1: mice was fed a normal nonpurified diet, composed of	Eight-week HFD increased endocannabinoid levels
	normal mice pellet; 2: mice was fed a HFD, containing	in all tissues except the liver and epididymal AT,
	21 wt% butter-fat and 0.15 wt% cholesterol; 3-5: mice,	and KO reduced anandamide and/or 2-AG levels in
	respectively, were fed a HFD with increasing doses of	all tissues but not in the liver, usually in a dependent
	KO 1.25, 2.5 or 5% wt of KO.	manner.

2-AG: 2-Arachidonilglycerol; AA: Arachidonic acid; AEA: Anendamide; AT: Adipose tissue; DHA: Docosahexaenoic acid; ECS: Endocannabinoid system; EPA: Eicosapentaenoic acid; FO: Fish oil; HFD: High fat diet; KO: Krill oil; LA: Linoleic acid; LC-PUFA: Longchain PUFA; LFD: Low fat diet; LKO: Fabp1 gene ablation; SO: Soyabean oil, STD: Standard/low-fat diet.

#### 3. Conclusion

HFD leads to increased endocannabinoids, increased endocannabinoids cause activation of CB1 receptors by activating ECS. This stimulates hypothalamus, including hedonic system and limbic system, leading to increased food intake; stimulates of adipogenesis in adipose tissue, decreases adiponectin production, increases lipogenesis and reduces fatty acid oxidation, so it increases risk of developing metabolic diseases such as insulin resistance, Type 2 Diabetes and cardiovascular diseases, particularly obesity.

Preventing obesity increasing prevalence and reaching epidemic proportions globally, is easier and less costly than treatment. For this reason, especially regulation of amount and pattern of fat in dietary pattern is an important requirement in prevention of obesity.

**Conflict of interest:** No conflict of interest was declared by the authors.

#### 4. References

- 1. Nguyen PH, Le TV, Kang HW, Chae J, Kim SK, Kwon KI *et al.* AMP-activated protein kinase (AMPK) activators from Myristica fragrans (nutmeg) and their anti-obesity effect. Bioorg Med Chem Lett. 2010; 20(14):4128-31.
- 2. Williams EP, Mesidor M, Winters K, Dubbert PM, Wyatt SB. Overweight and obesity: prevalence, consequences, and causes of a growing public health problem. Curr Obes Rep. 2015; 4(3):363-70.
- 3. Wilson PW, D'Agostino RB, Sullivan L, Parise H, Kannel WB. Overweight and obesity as determinants of cardiovascular risk: the Framingham experience. Arch Intern Med. 2002; 162(16):1867-72.
- 4. Mandviwala T, Khalid U, Deswal A. Obesity and cardiovascular disease: a risk factor or a risk marker? Curr Atheroscler Rep. 2016; 18(5):21.
- Laviea CJ, De Schuttera A, Partoa P, Jahangira E, Kokkinosb P, Ortegac FB et al. Obesity and prevalence of cardiovascular diseases and prognosis—the obesity paradox updated. Prog Cardiovasc Dis. 2016; 58(5):537-47.
- 6. Afshin A, Forouzanfar MH, Reitsma MB, Sur P, Estep K, Lee A *et al.* Health effects of overweight and obesity in 195 countries over 25 years. N Engl J Med. 2017; 377(1):13-27.
- 7. Kirkham TC. Endocannabinoids in the regulation of appetite and body weight. Behav Pharmacol. 2005; 16:297-313.
- 8. Bensaid M, Gary-Bobo M, Esclangon A, Maffrand JP, Le Fur G, Oury-Donat F *et al*. The cannabinoid CB1 receptor

- antagonist SR141716 increases Acrp30 mRNA expression in adipose tissue of obese fa/fa rats and in cultured adipocyte cells. Mol Pharmacol. 2003; 63(4):908-14.
- 9. Engeli S, Bohnke J, Feldpausch M, Gorzelniak K, Janke J, Batkai S *et al.* Activation of the peripheral endocannabinoid system in human obesity. Diabetes. 2005; 54(10):2838-43.
- 10. Kunos G, Batkai S. Novel physiologic functions of endocannabinoids as revealed through the use of mutant mice. Neurochem Res. 2001; 26(8-9):1015-21.
- 11. Scheen AJ, Van Gaal LG, Despres JP, Pi-Sunyer X, Golay A, Hanotin C. Rimonabant improves cardiometabolic risk profile in obese or overweight subjects: overview of RIO studies. Rev Med Suisse. 2006; 2(76):1916-23.
- 12. Despres JP, Golay A, Sjostrom L. Rimonabant in Obesity-Lipids Study Group. Effects of rimonabant on metabolic risk factors in overweight patients with dyslipidemia. N Engl J Med. 2005; 353(20):2121-34.
- 13. Van Gaal LF, Rissanen AM, Scheen AJ, Ziegler O, Rössner S. RIO-Europe Study Group. Effects of the cannabinoid-1 receptor blocker rimonabant on weight reduction and cardiovascular risk factors in overweight patients: 1-year experience from the RIO-Europe study. Lancet. 2005; 365(9468):1389-97.
- 14. Curioni C, Azara C, Capelli J. Rimonabant for Obesity. In Atta-ur-Rahman FRS. M. Iqbal Choudhary Editors. Anti-Obesity Drug Discovery and Development. 1th ed. Taiwan: Bentham Science. 2011, 131-49.
- 15. Di Marzo V. Endocannabinoids: synthesis and degradation. Rev Physiol Biochem Pharmacol. 2008; 160:1-24.
- 16. Jager G, Witkamp RF. The endocannabinoid system and appetite: relevance for food reward. Nutr Res Rev. 2014; 27(1):172-85.
- 17. Banni S, Di Marzo V. Effect of dietary fat on endocannabinoids and related mediators: consequences on energy homeostasis, inflammation and mood. Mol Nutr Food Res. 2010; 54(1):82-92.
- Pertwee RG, Howlett AC, Abood ME, Alexander SP, Di Marzo V, Elphick MR et al. International Union of Basic and Clinical Pharmacology. LXXIX. Cannabinoid receptors and their ligands: beyond CB<sub>1</sub> and CB<sub>2</sub>. Pharmacol Rev. 2010; 62(4):588-631.
- 19. Ramsden CE, Zamora D, Makriyannis A, Wood JT, Mann JD, Faurot KR. *et al.* Diet-induced changes in n-3-and n-6-derived endocannabinoids and reductions in headache pain and psychological distress. J Pain. 2015; 16(8):707-16.

- 20. Salem N Jr, Pawlosky R, Wegher B, Hibbeln J. *In vivo* conversion of linoleic acid to arachidonic acid in human adults. Prostaglandins Leukot Essent Fatty Acids. 1999; 60(5-6):407-10.
- Cravatt BF, Demarest K, Patricelli MP, Bracey MH, Giang DK, Martin BR. Supersensitivity to anandamide and enhanced endogenous cannabinoid signaling in mice lacking fatty acid amide hydrolase. Proc Natl Acad Sci USA. 2001; 98(16):9371-6.
- 22. Okamoto Y, Morishita J, Tsuboi K, Tonai T, Ueda N. Molecular characterization of a phospholipase D generating anandamide and its congeners. J Biol Chem. 2004; 279(7):5298-305.
- 23. Venance L, Stella N, Glowinski J, Giaume C. Mechanism involved in initiation and propagation of receptor-induced intercellular calcium signaling in cultured rat astrocytes. J Neurosci. 1997; 17(6):1981-92.
- 24. Tsutsumi T, Kobayashi T, Ueda H, Yamauchi E, Watanabe S, Okuyama H. Lysophosphoinositide-specific phospholipase C in rat brain synaptic plasmamembranes. Neurochem Res. 1994; 19(4):399-406.
- 25. Naughton SS, Mathai ML, Hryciw DH, McAinch AJ. Fatty acid modulation of the endocannabinoid system and the effect on food intake and metabolism. Int J Endocrinol. 2013; 2013: 361895.
- 26. Williams CM, Rogers PJ, Kirkham TC. Hyperphagia in pre-fed rats following oral delta9-THC. Physiol Behav. 1998; 65(2):343-6.
- 27. Bellocchio L, Lafenêtre P, Cannich A, Cota D, Puente N, Grandes P *et al.* Bimodal control of stimulated food intake by the endocannabinoid system. Nat Neurosci. 2010; 13(3):281-3.
- 28. Pagotto U, Marsicano G, Cota D, Lutz B, Pasquali R. The emerging role of the endocannabinoid system in endocrine regulation and energy balance. Endocr Rev. 2006; 27(1):73-100.
- Di Patrizio NV, Astarita G, Schwartz G, Li X, Piomelli D. Endocannabinoid signal in the gut controls dietary fat intake. Proc Natl Acad Sci USA. 2011; 108(31):12904-8.
- 30. Schneeberger M, Gomis R, Claret M. Hypothalamic and brainstem neuronal circuits controlling homeostatic energy balance. J Endocrinol. 2014; 220(2):T25-46.
- 31. Utoyama M, Akieda-Asai S, Koda S, Nunoi H, Date Y. Role of the neural pathway from hindbrain to hypothalamus in the regulation of energy homeostasis in rats. Neurosci Lett. 2016; 614:83-8.
- 32. Bermudez-Silva FJ, Cardinal P, Cota D. The role of the endocannabinoid system in the neuroendocrine regulation of energy balance. J Psychopharmacol. 2012; 26(1):114-24.
- 33. Kola B, Farkas I, Christ-Crain M, Wittmann G, Lolli F, Amin F *et al*. The orexigenic effect of ghrelin is mediated through central activation of the endogenous cannabinoid system. PLoS One. 2008; 3(3):e1797.
- 34. Burdyga G, de Lartigue G, Raybould HE, Morris R, Dimaline R, Varro A *et al.* Cholecystokinin regulates expression of Y2 receptors in vagal afferent neurons serving the stomach. J Neurosci. 2008; 28(45):11583-92.
- 35. Rigamonti AE, Piscitelli F, Aveta T, Agosti F, De Col A, Bini S *et al.* Anticipatory and consummatory effects of

- (hedonic) chocolate intake are associated with increased circulating levels of the orexigenic peptide ghrelin and endocannabinoids in obese adults. Food Nutr Res. 2015; 59:29678.
- 36. Izzo AA, Sharkey KA. Cannabinoids and the gut: new developments and emerging concepts. Pharmacol Ther. 2010; 126:21-38.
- Burdyga G, Varro A, Dimaline R, Thompson DG, Dockray GJ. Ghrelin receptors in rat and human nodose ganglia: putative role in regulating CB-1 and MCH receptor abundance. Am J Physiol Gastrointest Liver Physiol. 2006; 290(6):G1289-97.
- 38. Lenard NR, Berthoud HR. Central and peripheral regulation of food intake and physical activity: pathways and genes. Obesity. 2008; 16(3):S11-22.
- 39. Muller T, Demizieux L, Troy-Fioramonti S, Gresti J, Pais de Barros JP, Berger H *et al.* Overactivation of the endocannabinoid system alters the anti-lipolytic action of insulin in mouse adipose tissue. Am J Physiol Endocrinol Metab. 2017; 313(1):E26-36.
- 40. Jamshidi N, Taylor DA. Anandamide administration into the ventromedial hypothalamus stimulates appetite in rats. Br J Pharmacol. 2001; 134(6):1151-4.
- 41. Wenger T, Jamali KA, Juanéda C, Léonardelli J, Tramu G. Arachidonyl ethanolamide (anandamide) activates the parvocellular part of hypothalamic paraventricular nucleus. Biochem Biophys Res Commun. 1997; 237(3):724-8.
- 42. Kirkham TC, Williams CM, Fezza F, Di Marzo V. Endocannabinoid levels in rat limbic forebrain and hypothalamus in relation to fasting, feeding and satiation: stimulation of eating by 2-arachidonoyl glycerol. Br J Pharmacol. 2002; 136(4):550-7.
- 43. Hanlon EC, Baldo BA, Sadeghian K, Kelley AE. Increase in food intake or food seeking behavior induced by GABAergic, opioid, or dopaminergic stimulation of the nucleus accumbens: is it hunger? Psychopharmacology (Berl). 2004; 172(3):241-7.
- 44. Soria-Gomez E, Matias I, Rueda-Orozco PE, Cisneros M, Petrosino S, Navarro L et al. Pharmacological enhancement of the endocannabinoid system in the nucleus accumbens shell stimulates food intake and increases c-Fos expression in the hypothalamus. Br J Pharmacol. 2007; 151(7):1109-16.
- 45. Monteleone P, Fabrazzo M, Tortorella A, Martiadis V, Serritella C, Maj M. Circulating ghrelin is decreased in non-obese and obese women with binge eating disorder as well as in obese non-binge eating women, but not in patients with bulimia nervosa. Psychoneuroendocrinology. 2005; 30(3):243-50.
- 46. Bluher M, Engeli S, Kloting N, Berndt J, Fasshauer M, Batkai S *et al.* Dysregulation of the peripheral and adipose tissue endocannabinoid system in human abdominal obesity. Diabetes. 2006; 55:3053-60.
- 47. Frieling H, Albrecht H, Jedtberg S, Gozner A, Lenz B, Wilhelm J *et al.* Elevated cannabinoid 1 receptor mRNA is linked to eating disorder related behavior and attitudes in females with eating disorders. Psychoneuroendocrinology. 2009; 34(4):620-4.
- 48. Cota D, Marsicano G, Tschöp M, Grübler Y, Flachskamm

- C, Schubert M *et al*. The endogenous cannabinoid system affects energy balance via central orexigenic drive and peripheral lipogenesis. J Clin Invest. 2003; 112(3):423-31.
- Osei-Hyiaman D, De Petrillo M, Pacher P, Liu J, Radaeva S, Sándor Bátkai S *et al*. Endocannabinoid activation at hepatic CB1 receptors stimulates fatty acid synthesis and contributes to diet-induced obesity. J Clin Invest. 2005; 115(5):1298-305.
- 50. De Gottardi A, Spahr L, Ravier-Dall'Antonia F, Hadengue A. Cannabinoid receptor 1 and 2 agonists increase lipid accumulation in hepatocytes. Liver Int. 2010; 30(10):1482-9.
- 51. Tam J, Vemuri VK, Liu J, Bátkai S, Mukhopadhyay B, Godlewski G *et al.* Peripheral CB1 cannabinoid receptor blockade improves cardiometabolic risk in mouse models of obesity. J Clin Invest. 2010; 120(8):2953-66.
- 52. Pacher P, Bátkai S, Kunos G. The Endocannabinoid system as an emerging target of pharmacotherapy. Pharmacological rev. 2006; 58(3):389-462.
- 53. Kola B, Hubina E, Tucci SA, Kirkham TC, Garcia EA, Mitchell SE *et al.* Cannabinoids and ghrelin have both central and peripheral metabolic and cardiac effects via AMP-activated protein kinase. J Biol Chem. 2005; 280(26):25196-201.
- 54. Spoto B, Fezza F, Parlongo G, Battista N, Sgro' E, Gasperi V *et al.* Human adipose tissue binds and metabolizes the endocannabinoids anandamide and 2-arachidonoylglycerol. Biochimie. 2006; 88(12):1889-97.
- 55. Savva SC, Chadjigeorgiou C, Hatzis C, Kyriakakis M, Tsimbinos G, Tornaritis M *et al.* Association of adipose tissue arachidonic acid content with BMI and overweight status in children from Cyprus and Crete. Br J Nutr. 2004; 91(4):643-9.
- 56. Muccioli GG, Naslain D, Bäckhed F, Reigstad CS, Lambert DM, Delzenne NM *et al.* The endocannabinoid system links gut microbiota to adipogenesis. Mol Syst Biol. 2010; 6:392-407.
- 57. Osei-Hyiaman D, Liu J, Zhou L, Godlewski G, Harvey-White J, Jeong WI *et al.* Hepatic CB1 receptor is required for development of diet-induced steatosis, dyslipidemia, and insulin and leptin resistance in mice. J Clin Invest. 2008; 118(9):3160-9.
- 58. Matias I, Gonthier MP, Orlando P, Martiadis V, De Petrocellis L, Cervino C *et al.* Regulation, function, and dysregulation of endocannabinoids in models of adipose and beta-pancreatic cells and in obesity and hyperglycemia. J Clin Endocrinol Metab. 2006; 91(8):3171-80.
- 59. Pagano C, Rossato M, Vettor R. Endocannabinoids, adipose tissue and lipid metabolism. J Neuroendocrinol. 2008; 20(1):124-9.
- 60. Annuzzi G, Piscitelli F, Di Marino L, Patti L, Giacco R, Costabile G *et al.* Differential alterations of the concentrations of endocannabinoids and related lipids in the subcutaneous adipose tissue of obese diabetic patients. Lipids in Health and Disease. 2010; 9:43-51.
- 61. Liu Y, von Deneen KM, Kobeissy FH, Gold MS. Food addiction and obesity: evidence from bench to bedside. J Psychoactive Drugs. 2010; 42(2):133-45.

- 62. Merlo LJ, Klingman C, Malasanos TH, Silverstein JH. Exploration of food addiction in pediatric patients: A preliminary investigation. J Addict Med. 2009; 3(1):26-32.
- 63. Kohjima M, Sun Y, Chan L. Increased food intake leads to obesity and insulin resistance in the TG2576 alzheimer's disease mouse model. Endocrinology. 2010; 151(4):1532-40.
- 64. Cota D. Role of the endocannabinoid system in energy balance regulation and obesity. Front Horm Res. 2008; 36:135-45.
- 65. Di Marzo V, Matias I. Endocannabinoid control of food intake and energy balance. Nat Neurosci. 2005; 8(5):585-
- 66. Chentouf M, Dubois G, Jahannaut C, Castex F, Lajoix AD, Gross R et al. Excessive food intake, obesity and inflammation process in Zucker fa/fa rat pancreatic islets. PLoS One. 2011; 6(8):e22954.
- 67. Bellocchio L, Cervino C, Pasquali R, Pagotto U. The endocannabinoid system and energy metabolism. J Neuroendocrinol. 2008; 20(6):850-7.
- 68. Matias I, Gatta-Cherifi B, Tabarin A, Clark S, Leste-Lasserre T, Marsicano G *et al.* Endocannabinoids Measurement in Human Saliva as Potential Biomarker of Obesity. PLoS One. 2012; 7(7):e42399.
- 69. Eckardt K, Sell H, Taube A, Koenen M, Platzbecker B, Cramer A *et al.* Cannabinoid type 1 receptors in human skeletal muscle cells participate in the negative crosstalk between fat and muscle. Diabetologia. 2009; 52(4):664-74
- 70. Vettor R, Pagano C. The role of the endocannabinoid system in lipogenesis and fatty acid metabolism. Best Pract Res Clin Endocrinol Metab. 2009; 23(1):51-63.
- 71. Engeli S, Lehmann AC, Kaminski J, Haas V, Janke J, Zoerner AA *et al*. Influence of dietary fat intake on the endocannabinoid system in lean and obese subjects. Obesity (Silver Spring). 2014; 22(5):E70-6.
- 72. Alvheim AR, Torstensen BE, Lin YH, Lillefosse HH, Lock EJ, Madsen L *et al.* Dietary linoleic acid elevates endogenous 2-arachidonoylglycerol and anandamide in Atlantic salmon (Salmo salar L.) and mice, and induces weight gain and inflammation in mice. Br J Nutr. 2013; 109(8):1508-17.
- 73. Crespillo A, Suárez J, Bermúdez-Silva FJ, Rivera P, Vida M, Alonso M *et al.* Expression of the cannabinoid system in muscle: effects of a high-fat diet and CB1 receptor blockade. Biochem J. 2011; 433(1):175-85.
- 74. Alvheim AR, Malde MK, Osei-Hyiaman D, Lin YH, Pawlosky RJ, Madsen L *et al.* Dietary linoleic acid elevates endogenous 2-AG and anandamide and induces obesity. Obesity (Silver Spring). 2012; 20(10):1984-94.
- 75. Martin GG, Landrock D, Chung S, Dangott LJ, Seeger DR, Murphy EJ *et al*. Fabp1 gene ablation inhibits high-fat diet-induced increase in brain endocannabinoids. J Neurochem. 2017; 140(2):294-306.
- Batetta B, Griinari M, Carta G, Murru E, Ligresti A, Cordeddu L *et al.* Endocannabinoids may mediate the ability of (n-3) fatty acids to reduce ectopic fat and inflammatory mediators in obese Zucker rats. J Nutr. 2009; 139(8):1495-501.

- 77. Matias I, Petrosino S, Racioppi A, Capasso R, Izzo AA, Di Marzo V. Dysregulation of peripheral endocannabinoid levels in hyperglycemia and obesity: Effect of high fat diets. Mol Cell Endocrinol. 2008; 286(1-2 Suppl 1):S66-78.
- 78. Piscitelli F, Carta G, Bisogno T, Murru E, Cordeddu L, Berge K *et al.* Effect of dietary krill oil supplementation on the endocannabinoidome of metabolically relevant tissues from high-fat-fed mice. Nutr Metab (Lond). 2011; 8(1):51-68.